Pak. J. Pharm. 20-23 (1 & 2) 37-41, 2007-2010



Review Article
ISSN: 1019-956X

A REVIEW ON Carthamus oxycantha

Mansoor Ahmad¹*, Imran Waheed¹, Muhammad Khalil-ur-Rehman², Uzma Niaz² and Syed Saeed-ul-Hassan²

¹Faculty of Pharmacy, University of Karachi, Karachi, Pakistan ²College of Pharmacy, University of the Punjab, Allama Iqbal campus, Lahore-54000, Pakistan

ABSTRACT

South Asian Regions including India, Pakistan and Bangladesh are very rich in medicinal plants and have about 10% of world's plant diversity. The traditional systems of medicine have deep penetration in this region. A large proportion of the society here, purely or largely have to depend on these systems. The reason behind is the effectiveness of these traditional healing systems or is the socioeconomic status. These systems mainly rely on the natural sources or on the plants, herbs or the weeds. Among these medicinally important plants is *Carthamus oxycantha*, of Asteraceae family, a common weed and has demonstrated beneficial biological activities like wound healing and others.

Keywords: Carthamus oxycantha, Asteraceae, Linoleic acid

INTRODUCTION

Traditional use of medicines is recognized as a way to learn about potential future medicines. About 122 compounds currently used in medicine, are derived from "ethnomedical" plant sources. About 80% of these compounds are used in medicine as such as were used traditionally. Plants play vital role in the world economy. Whatever the field is the role of plant is worthwhile. Utilization of plants in alternative medicine is also a stance of plant role (Rehman et al., 2001; Fabricant and Farnsworth 2001). Many plant families yield medicinally important plants, Asteraceae is one of them. Many locally occurring species of Asteraceae are used as medicinal plants by various tribal and ethnic communities in Pakistan. Carthamus oxycantha is often occurring as weed in cultivated fields. Folk medicines indicate its use as antiinflammatory and wound healing properties. Its healing and/or counter irritant activity is also evident from studies conducted on the albino rabbits (Waheed, 2009).

Plant Family Asteraceae

Asteraceae is the largest family of dicotyledon Angiosperm. It includes more than 1000 genera and

about 10,000 species distributed over all parts of the world. It mostly contain herbs or under – shrubs, rarely trees or climbers, with leaves usually alternate or opposite and exstipulate. Flowers are crowded into heads (capitula), surrounded at the base by one or more whorls of free or united bracts (involucre), sometime the heads are compound and the partial heads few or one flowered; floral bracts entirely absent or reduced to scales or bristles on the flat, conical or rarely elongated or concave receptacle (Parker, 1918; Kashyap and Joshi, 1936).

Flowers all tubular (heads discoid), or the inner tubular and outer ringulate (head radiate), or all ligulate (head ligulate), all bisexual. Calyx – tube adnate to the ovary; limb are absent, or of scales, bristles or hairs. Corolla gamopetalous, epigynous, regular, either tubular, 4-5lobed and actinomorphic, or ligulate, or rarely bilabiate and zygomorphic; stamens 5, rarely 4, epipetalous, filaments free; anthers generally connate into a tube sheathing the style, rarely free, 2-celled, opening lengthwise, base of the cells often tailed or prolonged downwards; ovary inferior, one celled, ovule solitary, erect from the base, anatropous; style usually divided at the top into stigmatic arms. Fruit is an achene naked or

*Corresponding author's Address: Prof. Dr. Mansoor Ahmad, Faculty of Pharmacy, University of Karachi, Karachi, Pakistan. E-mail: herbalist53@yahoo.com

Ahmad et al

crowned by the persistent sessile or stipitate papus, beaked. Seeds often ex-albuminous. sometime cotyledons straight, plano-convex. It is the biggest plant family in subcontinental region. It is a source of a number of medicinal plants which are used in allopathic as well as in traditional system of medicine, practiced in different part of the world. However, many plants of the family are used as medicines by various tribal and ethnic communities in subcontinent. It is interesting to note that these people are using some of the species for the ailments which are not mentioned in any of the traditional systems of medicines (Parker, 1918; Kashyap and Joshi, 1936).

As regard the chemical constituents present in the members of this family, many authors have investigated a large number of species of the family. The chemotaxonomy of the family was outlined by Herout (Herout, 1966; Herout, 1970). Bacon and Edelman (1951) isolated carbohydrates, Quillet and Bourdu (1952) separated glucoside and Politis (1950) investigated chlogrogenic acid present in the subterranean organs of various species of this family. Large number of sesquiterpene lactones and their isomers from a number of species of this family have been isolated and characterized by many workers (Richard, 1965; Rybalko *et al.*, 1965; Kery *et al.*, 1987; Milosavljevic, 1995; Milosavljeric *et al.*, 1999; DaCosta *et al.*, 2005).

Flavones and flavonoid compounds have been isolated from many species of this family (Bandyukova, 1968; Kaneta *et al.*, 1978). The terpenes and their derivatives were isolated and characterized (Pyrek and Baranowska, 1973; Pyrek, 1977; Bader *et al.*, 1990; Alvarenga *et al.*, 1995). Aromatic polyacetylenes, thiophenes, coumarins derivatives and pyrrolizidine alkaloids have also been isolated and characterized (Bauthrope and Brown, 1989; Paiz *et al.*, 1989; Balz *et al.*, 1993; Konovalov, 1996; Liddell, 1998). A number of trace elements present in aqueous extract of different species of Asteraceae were detected and isolated (Yin *et al.*, 1994).

The polysaccharides isolated from medicinal plants of this family stimulate the healing process of gastric ulcer in rats. Anticancer, cytotoxic, anti-microbial and toxicity of terpenoids and thiophenes present in a number of species of this family were investigated. Inhibitors of α glucosidase for treatment of obesity and diabetes and as antiaging cosmetics, bath cosmetic compounds and health food components for blood circulation disorders were also isolated from different species of this family. Further the anti-tumor extract from different plants was also evaluated (Lavrenova, and Chernov, 1983; Hudson *et al.*, 1993; Villarreal *et al.*, 1994; Pogrebnyak *et al.*, 1998; Jahodar, and Klecakova, 1999). The plant induced contact dermatitis, contact hypersensitivity to a perfume material, photosensitivity due to sesquiterpene lactones and the applications of hypo-irritatitve and hypo-allergic material obtained from a number of Asteraceae plants were evaluated by many works. Large number of natural insecticides, synergistic herbicides and nematocidal principles from different plants of Asteraceae have been isolated, characterized and are used in agriculture practices (Gommer, 1973; Mitchell *et al.*, 1974; Thune and Solberg, 1980; Okabe, 1994; Gonzalez-Coloma *et al.*, 2002; Iwata, 2002).

Genus – Carthamus Bieb. General botanical description

Puberulous, stem and braches while – leaves oblong or oblong lanceolate, lower shortly spinulose-toothed, upper half-amplexicaul very spinous-outer involucral bracts exceeding the head white below the contracted portion green above it with yellow spines – Flowers orange yellow – Achenes abouoid 4 – angled smooth shining truncate at the top with 4 bosses (Kirtikar and Basu, 1985).

Carthamus oxycantha Bieb.

Vernacular Names Arabic: Qartam Enlish: Jeweled Distaff thistle, wild safflower Punajabi: Kandiari Pushto: Kunzalay Urdu.: Pholi

Classification

Kingdom	Plantae – Plants
Subkingdom	Tracheobionta – Vascular Plants
Superdivision	Spermatophyta – Seed Plants
Division	Magnoliopsida – Flowering Plants
Class	Magnoliopsida – Dicotyledons
Subclass	Asteridae
Order	Asterales
Family	Asteraceae – Aster Family
Genus	Carthamus L.
Species	Carthamus oxycantha Bieb.

Distribution

The plant is mostly found in dry, open places, plains and mountains. It is distributed in Afghanistan, Azerbaijan, India, Iran, Iraq, Kyrgyzstan, Pakistan, Tajikistan and Turkmenistan. It is a common weed along the boundaries of wheat field (Kashyap and Joshi, 1936; Nadkarni, 1976; Chopra *et al.*, 1982).

Description

Plant is a stout, strongly armed, generally much branched, puberulous herb, 1- 1 ¹/₂ feet tall, stem and branches white. Leaves sessile, 1-5 inches long, oblong or oblong-lanceolate; lower often pinnatified, shortly

spinulose-toothed; upper $\frac{1}{2}$ - amplexicaul, very spinous. Heads $\frac{3}{4}$ - $\frac{1}{2}$ inche in diameter; outer involucral bracts exceeding the head, white below the contracted portion, green above it, with yellow spines. Flowers are orange yellow. Achenes obovoid 4- angled smooth shinning truncate at the top with 4 bosses, pappus 0 (Kirtikar and Basu, 1985). Found abundantly in the fields after the crops have been harvested. It flowers in March – June (Kashyap and Joshi, 1936).

Chemical constituents

Carthamus oxycantha seeds yields two types of oils: oleic oil and linoleic oil. Fatty acid oil composition of oleic oil is, palmitic acid 5-6%, stearic acid 1.5 -2%, oleic acid 74-80%, linoleic acid 13-18% and traces of linoleic acid and longer chain fatty acids. The fatty acid composition of linoleic oil, palmitic acid 5-8%, stearic acid 2-3%, oleic acid 8-30%, linoleic acid 67-89% and also traces of linoleic acid and longer chain fatty acids. Carthamus oxycantha fruit also contains proteins 20-25%, hull 60%, residual fat 2-15%. Flowers of Carthamus oxycantha contain two major pigments, the water soluble, yellow carthamidin and the formally important dye carthamin, flavonone which is orange red (Fernandez-Martinez et al., 1993; Anjani, 2005). Flowers also contain 0.3-0.6% carthamin. Flavonoids, glycosides, sterols and serotonin derivatives have been identified from flowers and seeds (Firestone, 1999). Two new glycosides, 2-O-methylglucopyranosyl-carthamoside and beta-D-fructofuranosyl carthamoside, along with the known compound 3', 4', 5, 7-tetrahydroxyflavanone have been isolated from Carthamus oxyacantha using recycling preparative HPLC. The structures of these compounds were established by mass spectrometric and extensive spectroscopic analysis (Hassan et al., 2010). Structures of the important chemical constitutes of the plant are shown in Figure I.

Pharmacological actions and medicinal uses

Young leaves of *Carthamus oxycantha* are used as a vegetable, whereas seeds are used in cooking. The fruit is used as bird feed. *Carthamus oxycantha* herbage is valuable as green fodder in many countries. The straw is also used as fodder. Flowers are used to treat cerebral thrombosis, male infertility, rheumatism and bronchitis (Fernandez-Martinez *et al.*, 1993; Anjani, 2005). It also induces labour and is used as a tonic tea to invigorate heart and blood circulation. *Carthamus oxycantha* based medicines also show beneficial effect on pain and swelling associated with trauma. Flowers of this species are also used to treat jaundice, while the seeds are considered as laxative. The sap is believed to reduce salivation. The oil is applied to treat scabies. The edible

oil extracted from the seed is now the main product of *Carthamus oxycantha*. The oil is suitable for paint production. It is used mainly in cooking and for making salad dressings and margarine.



Figure I Structure of compounds in *Carthamus* oxycantha

Carthamus oxycantha had been used traditionally to make roghan wax used in the batik industry (Firestone, 1999). *Carthamus oxycantha* had long been grown for the dye extracted from the flowers. Depending on the dyeing procedure and the addition of other colourants and mordants, it imparts a yellow, red, brown or purple colour to cloth. However, dyes were still produced on a small scale for traditional and religious purposes. *Carthamus oxycantha* was a substitute or adulterant for true saffron as a natural food colourant. Flowers of this species were commonly mixed with rice, bread, pickles and other food to give them an attractive orange colour.

Ahmad et al

The seed cake was used as animal feed. The cake from undecorticated seeds containing matairesinol-glucoside was only suitable for ruminants. *Carthamus oxycantha* meal and flour from decorticate seed were used in the production of high-protein human diet supplements (Anjani, 2005).

CONCLUSION

Plants provide enormous reservoir of medicinal candidates. Though it is difficult to precisely study the potential natural candidates or active biomolecules due to variability and ecological issues but it helps to utilize the medicinal benefits of these plants. Screening of these agents from such plants as *Carthamus oxycantha* can lead to achieve such benefits and at national level can contribute in revenue generation Valuable plant constituents can be a modern medicinal products or their key biosynthetic precursor.

REFERENCES

- Alvarenga, S., Vestri, A. and Rodrigues, G. Do vale. (1995). Natural product substitution pattern and skeletal recognition method. *Natural Product Letter* 7, 133-140. *Chem. Abst.* 125, 222207m, (1996).
- Anjani, (2005). Genetic variability and character association inwild safflower (*Carthamus oxycantha*). *Indian Journal of Agricultural Science*. 75(8), 516-518.
- Bacon, J.S.D. and Edelman, J. (1951). Carbohydrates of the Jerusalem Artichoke and other Compositae. *Journal of Biochemistry* 48, 114-126. *Chem. Abst.* 45, 5242b, (1951).
- Bader, G., Hiller, K., Ehwald, R., Guempel, C. and Rathgen, K. (1990). Purification of pharmaceutical triterpene glycosides. DD 276, 287 Co71115/24), 21 Feb 1990, Appl. 320, 864, 18 Oct. 1988. *Chem. Abst.* 114, p3111b, (1991).
- Balz, F., Tower, G. and Neil, H. (1993). Naturally occurring C₁₃, dithiacyclo-hexadienes and thiophenes from the Asteraceae. *Methods in Plant Biochemistry*. 8, 551-572. *Chem. Abst.* 119, 265857s, (1993).
- Bandyukova, V.A. (1968). Distribution of flavonoids in some families of higher plants II. Family Compositae. *Rast. Resur.* 4, 429-441. *Chem. Abst.* 70, 17567h, (1969).
- Bauthrope, D.V. and Brown, G.D. (1989). Two unexpected coumarin derivatives from tissue cultures of Compositae species. *Phytochemistry*. 28, 3003-3007. *Chem. Abst.* 112, 135979c, (1990).
- Chopra, R. N., Chopra, I. C., Handa, K. L. and Kapur, L. D. (1982). Chopra's Indigenous Drugs of India. (2nd ed.). Academic Publishers, Calcutta, New Delhi, pp. 505-506.

- DaCosta, F.B., Terfloth, L. and Gastciger, J. (2005). Sesquiterpene lactone-based classification of three Asteraceae tribes; a study based on self- organizing neural networks applied to chemosystematics. *Phytochemistry*. 66, 345-353. *Chem. Abst.* 142, 333066k, (2005).
- Fabricant, D.S., Farnsworth N.R. (2001). The value of plants used in traditional medicine for drug discovery. *Environmental Health Perspectives*. 109, (1) 69–75.
- Fernandez-Martinez, M., Del-Rio, M. and de Haro, A. (1993). Survey of safflower *(Carthamus oxycantha L.)* germplasm for variants in fatty acid composition and other seed characters. *Euphytica*. 69,115-122.
- Firestone, D. (1999). Physical and chemical characteristics of oils, fats, and waxes. AOCS Press, Champaign, United States. pp 152-153.
- Gommer, F.J. (1973). Nematocidal principles in Compositae. Neth. Meded. Landbouw-hogesch. Wageninger 17, 71-73. Chem. Abst. 81, 59251h, (1974).
- Gonzalez-Coloma, A., Reina, M., Gutienez, C. and Fraga, B. M. (2002). Natural insecticides: structure activity relationships-a case study. *Studies of Natural Product Chemistry.* 26, 849-879. *Chem. Abst.* 137, 212230x, (2002).
- Hassan, Z., Ahmed, V.U., Hussain Z.J., Zahoor A, Siddiqui, I.N., Rasool Z.M.. (2010). Two new carthamosides from Carthamus oxycantha. *Natural Product Communication*. 5(3), 419-422.
- Herout, V. (1966). Chemotaxonomy of Compositae. Herba Hungarica., 5, 95-96. Chem. Abst. 68, 36724g, (1968).
- Herout, V. (1970). Chemotaxonomy of Compositae (Astreraceae). *Pharmacognosy and Phytochemistry International Congress.* 1, 93-110. *Chem. Abst.* 76, 124092v, (1972).
- Hudson, J.B., Graham, E.A., Rossi, R., Carpita, A., Neri, D. and Towers, G.H.N. (1993). Biological activities of terthiophenes and polyynes from the Asteraceae. *Planta Medica*. 59, 447-50. *Chem. Abst.* 120, 45283g, (1994).
- Waheed, I. (2009). Irritant contact dermatitic study of *Carthamus oxycantha* Bieb. M. Phil thesis submitted, College of Pharmacy, University of the Punjab, Lahore-Pakistan.
- Iwata, M. (2002). Plant-induced contact dermatitis. Arerugi, Men'eki. 9, 660-664. Chem. Abst. 138, 84609j, (2003).
- Jahodar, L. and Klecakova, J, (1999). Toxicity of Asteraceae with emphasis on pharmaceutically important species. *Chemicke Listy.* 93, 320-326. *Chem. Abst.* 131, 140530w, (1999).

- Kaneta, M., Hikichi, H., Endo, S. and Sugiyama, N. (1978). Identification of flavonoids in sixteen Compositae species. *Agricultural and Biological Chemistry*. 42, 475-477. *Chem. Abst.* 88, 186096f, (1978).
- Kashyap, S.R. and Joshi, A.C. (1936). Compositae. In: *Lahore District Flora*, The University of the Punjab, Lahore., pp. 139-140,149.
- Kery, A., Turiak, G., Zambo, I. and Tetenyi, P. (1987). Comparison of different separation techniques on the biologically active sesquiterpene lactones. *Acta Pharmacutica Hungarica*. 57, 228-238. *Chem. Abst.* 108, 62304j, (1988).
- Kirtikar, K.R. and Basu, B.D. (1985). *Indian Medicinal Plants* (2nd Edn.), Vol.1, International book distributors India, p.838-839.
- Konovalov, D.A. (1996). Aromatic polyacetylene compounds of the family Asteraccae and their taxonomic importance. *Rast. Resur.* 32, 84-98. *Chem. Abst.* 127, 31499q, (1997).
- Lavrenova, G.Yu. and Chernov, I.P. (1983). Comparative study of the effects of some plants polysaccharides on the inflammatory process and on regeneration in chronic gastric ulcer. *Farmacol Toksikol.* 46, 85-9. *Chem. Abst.* 99, 187261v, (1983).
- Liddell, J.R. (1998). Pyrrolizidine alkaloids. *Natural Product Reports.* 15, 363-370. *Chem. Abst.* 129, 260597d, (1998).
- Milosavljevic, S. (1995). Sesquiterpene lactones from wild Asteraceae in Yugoslavia. *ARH Farm.* 45, 199-206. *Chem. Abst.* 124, 140885w, (1996).
- Milosavljeric, S., Bulatovie, V. and Stefanovic, M. (1999). Sesquiterpene Lactones from wild growing plant families Asteraceae and Apiaceae. *Journal of Serbian Chemical Society*. 64, 397-442. *Chem. Abst.* 131, 240349n, (1999).
- Mitchell, J., John, C. and Epstein, W.L. (1974). Contact hypersensitivity to a perfume material, costus absolute.
 Role of sesquiterpene lactones. *Archives of Dermatology*. 110, 871-873. *Chem. Abst.* 82, 1336943u, (1975).
- Nadkarni, K. M. (1976). Indian Materia Medica. Popular parakashan Pvt. Ltd. pp. 469-470.
- Okabe, M. (1994). The application of the medicinal plants to the hypo-irritative and hypo-allergic cosmetics, mainly about the Labiatae at Compositae plants. *Fragrance Journal.* 22, 74-75. *Chem. Abst.* 121, 186732k, (1994).
- Paiz, L., Lopez, I. and Rodriguez, E. (1989). Chemistry and distribution bioactive polyacetylenes in Asteracae. *Revista Latinoamericana de Quimica*. 20, 120-125. *Chem. Abst.* 112, 135959w, (1990).

- Parker, R. N. (1918). Compositae. In: A Forest Flora for the Punjab with Hazara and Delhi, Printed by the Superintendent, Government Printing, Punjab, Lahore., pp. 286-287.
- Pogrebnyak, A. V., Poroikov, V.V., Starykh, V.V. and Konovalor, DA. (1998). Computer forecast of anticancer activity of sesquiterpence lactones discovered in representatives of Asteraceae. *Rast. Resur.* 34, 61-64. *Chem. Abst.* 129, 225333t, (1998).
- Politis, J. (1950). Cytological investigations on the mode of formation chlorogenic acid in the Compositae. *Proceedings of International Botany Congress.* 7, 367-368. *Chem. Abst.* 48, 12251e, (1954).
- Pyrek, J.S. and Baranowska, E. (1973). Faradiol and arnidiol. Revision the structure. *Tetrahedron Letters*. 11, 809-810. *Chem. Abst.* 79, 18885h, (1973).
- Pyrek, J. St. (1977). Terpenes of Compositae plants. Part VI. Faradiol and arnidiol, revision of their structure. *Roczniki Chemii* 51, 2331-2341. *Chem. Abst.* 88, 191151z, (1978).
- Quillet, M. and Bourdu, R. (1952). Glucides stored irrsuoterrancean orga of Compositae at the beginning of the rest period. *Comptes Rendus*. 234, 1079-1081.
- Rehman, S., Hasnat, A., Hasan, C. M., Rashid, M. A and Ilias, M. (2001). Pharmacological Evaluation of Bangladeshi medicinal Plan - A Review. *Pharmacol. Biol.* 37, 202-207.
- Richard, J. T. (1965). Sesquiterpenoid lactones of Compositae. I. Coronopilin, II. Coronopilic acid III. Chamissionin. Order No.65-4872, 256 pp. *Dissertation Abstr.* 25.6244-6245. *Chem. Abst.* 63, 8414b, (1965).
- Rybalko, K. S., Perel'son, M. E., Shretee, M. I., Vlasov, I. A., Gubanor, M. G., Pimenov, R. E., Pimenova, N. P., Novasel, T. and Serebryakaova, A.A. (1965).
 Sesquiterpene lactones in plants of the Compositae. *Aptechn. Delo* 14, 37-41. *Chem. Abst.* 64, 3955e, (1966).
- Thune, P.O. and Solberg, Y. J. (1980). Photosensitivity and allergy to aromatic lichen acids, compositae oleoresins and other plant substances. *Contact Dermatitis.* 6; 64-71. *Chem. Abst.* 93, 63143q, (1980).
- Villarreal, M. L., Alvarez, L., Alonso, D., Navarro, V., Garcia, P. and Delgado, G. (1994). Cytotoxic and antimicrobial screening of selected termpenoids from Asteraceae species. *Journal of Ethnopharmacology*. 42, 25-29. *Chem. Abst.* 121, 124717a, (1994).
- Yin, C., Peng, L., Chong, Li. and Changcheng, Z. (1994). Studies on trace elements of some traditional Chinese medicine in Compositae from Yunnan. *Xuebao Ziran Kexue Ban.* 16, 63-67. *Chem. Abst.* 123, 280822c, (1995).