# *In vitro* biological control of *Fusarium oxysporum*causing wilt in *Capsicum annuum*.

## Irfan Yousaf Sahi, A.N. Khalid

Department of Botany, University of the Punjab, Lahore, 54590, Pakistan. \*E-mail: irfan\_sahi@pap.gov.pk

## Abstract

Five species of *Trichoderma* viz., *Trichoderma Viride*, *T. harzianum*, *T. Koningii*, *T. aureoviride* and *T. pseudokoningii* were evaluated for their *in vitro* antagonistic potential against *Fusarium oxysporum*, the cause of wilt disease in sweet peppers (*Capsicum annum*). Among the *Trichoderma* species *T. viride* showed the best performance *in vitro* biological control of *Fusarium oxysporum* followed by *T. harzianum*, *T. aureoviride*, *T. koningii* and *T. pseudokoningii*, respectively, resulting in 62, 36, 24, 18 and 6% reduction in colony growth of the test pathogenic fungus respectively.

Keywords: *Capsicum annuum*, Biological control, *Fusarium oxysporum*, Sweet pepper, *Trichoderma* spp.

# Introduction

Sweet pepper (*Capsicum annuum* L.) belongs to the family solanaceae, which is an important group of vegetables cultivated extensively in Pakistan and also widely cultivated in almost every country of the world. Sweet pepper is a summer crop and its total area under cultivation in Pakistan is about 91800 hectare, with total annual production of 115,000 tonnes. It thrives best in warm climate, where frost is not a problem during growing seasons. In general, it requires temperature ranging 25-35 °C. (Govt. of Pak., 2001).

Fusarium oxysporum Schlecht is associated with wilt disease of sweet pepper (Mushtaq and Hashmi, 1997). Fusarium wilt is the most important disease caused by F. oxysporum in sweet pepper plants. Fusarium wilt first appears as slight vein clearing on the outer portion of the younger leaves, followed by epinasty (downward drooping) of the older leaves. At the seedling stage, plants infected by F. oxysporum may wilt and die soon after symptoms appear. In older plants, vein clearing and leaf epinasty are often followed by stunting, yellowing of the lower leaves, formation of adventitious roots, wilting of leaves and young stems, defoliation, marginal necrosis of remaining leaves, and finally death of the entire plant. Browning of the vascular tissue is an strong evidence of Fusarium wilt in sweet pepper (Agrios, 1988).

Healthy plants can become infected by F. oxysporum if the soil in which they are growing is contaminated with the fungus. The fungus can invade a plant with its sporangial germ tube or mycelium by invading the plant's roots. The roots can be infected directly through the root tips, through wounds in the roots, or at the formation point of lateral roots. Once inside the plant, the mycelium grows through the root cortex intercellulary. When the mycelium reaches the xylem, it invades the vessels through the xylem's pits. (Agrios, 1988).

Due to the growth of the fungus within the plant's vascular tissue, the plant's water supply is greatly affected. This lack of water induces the leaves' stomata to close, the leaves wilt, and the plant eventually dies. It is at this point that the fungus invades the plant's parenchymatous tissue, until it finally reaches the surface of the dead tissue, where it sporulates abundantly. The resulting spores which on their turn act as new inoculum for further spread of the fungus (Agrios, 1988).

Currently effective means of controlling *F.oxysporum* include: disinfestation of the soil and planting material with fungicidal chemicals, crop rotation with non-hosts of the fungus, or by using resistant cultivars (Jones *et al.*, 1982; Agrios, 1988; Smith *et al.*, 1988)

Use of environmentally friendly biological control agents can more affectively control the soilborne phytopathogens. (Nam *et al.*, 1988; Park ,1989; Saleem *et al.*, 2000).

From several studies, it has been confirmed that *Trichoderma spp.* have antagonistic and biologically control potential against a diversity of soil borne pathogens.(Grondona *et al.*, 1997; Hanson and Howell, 2004; Bajwa *et al.*, 2004).

Present study has been carried out to biologically control wilt disease in sweet pepper by using *Trichoderma spp*. as biocontrol agents.

# **Materials and Methods**

#### **Isolation of the Pathogen**

Root samples of 10 sweet pepper plants infected with wilting were collected from vegetable garden, Quaid–e–Azam campus, University of the Punjab, Lahore, Pakistan during June 2005. The root samples were cut into small pieces up to 1.5 cm length and surface sterilized by 15 %  $H_2O_2$  for 30-45 seconds. And then rinsed with distilled water for three times. These surface sterilized roots were placed onto 2% Malt Extract Agar (MEA) medium in Petri plates and incubated at 25°C After 6 days the fungal isolates appearing on the root pieces were identified and transferred to 2% MEA medium Petri plates for purification.

#### *In vitro* Biological Control

Cultures of F.oxysporum, isolated from roots of diseased ( wilted) sweet pepper plants, were maintained on 2% malt extract agar (MEA) medium . Cultures of various antagonistic fungi Trichoderma Viride Pers. ExGray, T.harzianum Rifai, T.Koningii Oudem, T.aureoviride Rifai, T.pseudokoningii Rifai were obtained from First Fungal Culture Bank of Pakistan, Department of Mycology and Plant Pathology, University of the Punjab, Lahore, Pakistan and maintained on 2% MEA medium. The antagonistic effect of various test fungal species in inhibiting the growth of Fusarium oxysporum was studied by Well method. For this purpose 6mm (0.6cm) diameter plugs of F. oxysporum and various antagonists were taken with the help of sterilized cork borer and placed at the opposite sides of the Petri plates of 9cm diameter having 2% MEA medium. After inoculation plates were incubated at 25°C. Petriplates with only F.oxysporum served as control. Each treatment was replicated for three times. Data on mycelial growth in terms of colony diameter of the pathogenic fungus were taken after 5 days of inoculation. Data was analyzed by using  $X^2$  test and level of significant was exact (P=0.05) followed by Duncun Multiple Range (DMR) test (Steel and Torrie, 1980) using computer software programme SPSS 10.

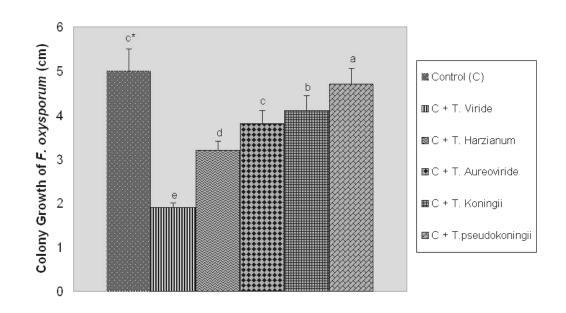
## **Results and Discussion**

Effect of Trichoderma species on in vitro growth of F. oxysporum is shown in fig 1& 2. Data recorded after 5 days of inoculation of biological control plates showed much more influence of antagonistic Trichoderma species over the colony growth of *F.oxysporum*. Among the Trichoderma species, T.viride showed the best performance where the colony growth of the pathogenic fungus F.oxysporum was 1.9cm as compared to 5cm in control treatment (Fig. 1). Trichoderma viride, antagonistic fungus, suppressed the colony growth of pathogen by 62% (Fig. 2). T.viride is also known to biologically control chestnut blight caused by Cryphonecteria parasitica in chestnut (Arisan-Atac et al., 1995). T.harzianum followed T.viride in performance and decreased colony growth of F.oxysporum by 36%. Rest of the three species of Trichoderma viz. T.aureoviride, T.koningii and T.pseudokoningii reduced the colony growth of F.oxysporum 24, 18 and 6% respectively. Effect of all the tested Trichoderma species except T.pseudokoningii was statistically significant against the pathogenic fungus F.oxysporum. Inoculation of sweet pepper (Capsicum annuum) plants with the tomato wilt pathogen, Fusarium oxysporum f.sp. lycopersici (FOL), partially protected pepper plants from subsequent infection with Phytophthora capsici, with no damage to plant.( Diaz et al., 2005). It has been observed that antagonistic fungi are specific in their antagonistic activity against specific fungi (Saleem et al, 2000). Trichoderma species are capable of producing extracellular lytic enzymes that are responsible for their antagonistic activity( Elad et al., 1982). Brasier (1975) reported that volatiles released by Trichoderma species reduced the growth of Phytophthora spp.

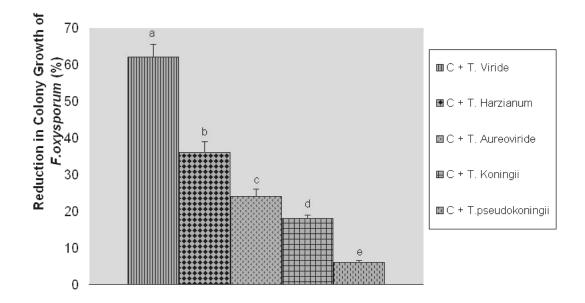
From the present study it is concluded that *Trichoderma* species have the potential to suppress the colony growth of *F.oxysporum* which is the pathogen of wilt in sweet peppers. Further studies are required to apply this technique of biocontrol in field of sweet pepper.

#### Acknowledgement

We are thankful to First Fungal Culture Bank of Pakistan for providing *Trichoderma spp*. Cultures. We are also thankful to Prof. Dr. S.H. Iqbal, Dr. Tehmina Anjum, Mr. Abdur Rehman and Miss Najum Sahar for their kind help in our research work.



**Fig. 1:** Colony growth of *F. oxysporum* in control (c) and in the presence of various antagonistic *Trichoderma* species. Bars with different letters at their tops show significant difference (P=0.05) as determined by DMR test.



**Fig. 2:** Percentage reduction in Colony growth of *F.oxysporum* due to various antagonistic *Trichoderma* species. Bars with different letters at their tops show significant difference (P=0.05) as determined by DMR test.

#### References

- Agrios GN, 1988. Plant Pathology, 3rd. ed. Academic Press, Inc.: New York. 803.
- Arisan-Atac I, Heidenreich E, Kubicek CP, 1995. Randomly amplified polymorphic

DNA fingerprinting identifies subgroups of *Trichoderma viride* and other *Trichoderma* sp. capable of chestnut blight biocontrol. FEMS Microbiol Lett. **126**: 249–256.

Bajwa R, Mukhtar I, Anjum T, 2004. In vitro biological control of Fusarium solani-

cause of wilt in *Dalbergia sissoo Roxb*. Mycopath, **2**(1): 11-14

- Brasier CM, 1975. Stimulation of sex organ formation of *Phytophthora* by antagonistic species of *Trichoderma*. I. The effect *in vitro*. *New Phyto.*, **74**:183-194
- Diaz J, Silvar C, Varela MM, Merino F, 2005. *Fusarium* confers protection against several mycelial pathogens of pepper plants, *Plant Pathol.* **54**:773-780
- Elad Y, Chet I, Katan J, 1982. Degradation of plant pathogenic fungi by *Trichoderma harzianum. Can. J. Microbiol.*, **28**: 719-725.
- Govt. of Pak, 2001. Agric. Statistics of Pak. Ministry of Food and Agriculture, Islambad. P.71-72.
- Grondona I, Hermosa R, Tejada M, Gomis MD, Mateos PF, Bridge PD, Monte E, Garcia-Acha I, 1997 Physiological and biochemical characterization of *Trichoderma harzianum*, a biological control agent against soilborne fungal plant pathogens. *Appl Environ Microbiol*. 63:3189–3198
- Hanson LE, Howell CR, 2004. Elicitors of plant defense responses from biological control strains of *Trichoderma virens*. *Phytopathology.*, **94**: 171 – 176.

- Jones JP, Jones JB, Miller W, 1982. *Fusarium* wilt on tomato. Fla. Dept. Agric. & Consumer Serv., Div. of Plant Industry. Plant Pathology Circular No. 237.
- Mushtaq M, Hashmi MH, 1997. Fungi associated with wilt disease of *Capsicum* in Sindh, Pakistan. *Pak. J. Bot.*, **29**(2): 217-222
- Nam CG, Jee HJ, Kim CH, 1988. Studies on biological control of Phytophthorra blight of red pepper. *Korean J. Plant Pathol.*, 4(4): 313-318
- Park, JH, 1989. Biological control of *Phytophthora* crown rot and root rot of greenhouse pepper with *Trichoderma harzianum* and *Enterobactor agglomerans* by improved methods of application. Korean J. Pl. and Patholol., **5(1):1**-12.
- Saleem A, Hamid K, Tariq AH, Jamil FF, 2000. Chemical control of root and collar rot of chillies. *Pak. J. Phytopath.*, **12**(1): 1-5
- Smith IM, Dunez J, Phillips DH, Lelliott RA, Archer SA, 1988. European handbook of plant diseases. Blackwell Scientific Publications. Oxford: 583
- Steel RGD, Torrie JH, 1980. Principles and procedures of statistics. A Biometrical approach. 2<sup>nd</sup> edition. McGraw Hill Book Co. Inc. New York, USA.