



Research Article

Bilateral Orbital Myiasis Due to *Wohlfahrtia magnifica* and its Treatment in a Dog

Amir Masoud Jafari-Nozad^{1*}, Keyvan Samadi², Kamran Akbarzadeh³, Hasan Bakhshi^{4*}, Kourosh Arzamani⁴ and Amirsajad Jafari⁵

¹Student Research Committee, Birjand University of Medical Sciences, Birjand, Iran.

²Oxygen Veterinary Clinic, Karaj, Iran.

³Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran.

⁴Vector-Borne Diseases Research Center, North Khorasan University of Medical Sciences, Bojnurd, Iran.

⁵Department of Basic Sciences, School of Veterinary Medicine, Shiraz University, Shiraz, Iran.

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Authors' Contributions

ASJ and HB conceived the presented idea. KS and AMJ isolated the samples. KS treated the case. KAA and KOA performed species identification. AMJ, ASJ and HB wrote the manuscript. ASJ and HB supervised the whole process.

Keywords

Dog, *Wohlfahrtia magnifica*, Myiasis, Treatment



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Abstract | Myiasis is described as infection and infestation by dipterous larvae in organs or different tissues of humans and animals. Ocular myiasis is defined as eye or periorbital tissue infestation by dipterous larvae. Larval stages of the *Wohlfahrtia magnifica* (Diptera: Sarcophagidae) parasitize many warm-blooded vertebrates resulting in a severe myiasis of mucosal membranes and wound lesions. In the present survey, a rare case of bilateral ocular myiasis due to *W. magnifica* in an ownerless dog in Iran is described. Physical examinations showed massive larvae over the eyes, unpleasant smell, edema and erythema, ulcers and necrotic tissues. Larvae were removed and were identified under the stereomicroscope with valid morphological keys. Furthermore, the treatment was carried out after removing the larvae, washing, using Oxytetracycline spray and topical permethrin, followed by the antibiotic therapy. The present report suggests that mechanical elimination of the larvae, subsequent washing of the wound with diluted Povidone-iodine, disinfection of the surrounding with permethrin and antibiotic therapy alongside anti-inflammatory drugs are effective in the treatment of orbital myiasis in dogs.

Novelty Statement | This study deals with a novel case of bilateral orbital myiasis in a dog and the treatment protocol with antiseptics and anti-inflammatory drugs.

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Introduction

Tick bites, surgical wounds, fight wounds, and perforations are the main lesions where flies prefer to

lay eggs. Myiasis is described as infection and infestation by fly larvae (maggot) in the organs or tissues of animals and humans. Myiasis may result in organ disfigurements, unpleasant smell, anorexia, and weight loss. In severe cases, toxemia and sepsis caused by secondary bacterial infection are possible (Najjari *et al.*, 2014; Nouri and Salehi, 2020). Based on the target organ, the myiasis is categorized as cutaneous, ocular, urogenital, nasal, auricular, and

Corresponding Author: Amir Masoud Jafari-Nozad and Hasan Bakhshi

a.sajad.jafari@gmail.com, hbakhshi89@gmail.com

so on (Scholl *et al.*, 2019). Ocular myiasis is defined as eye or periorbital tissue infestation by the maggots. Depending on the affected area of the eye, ocular myiasis is categorized into three groups: Ophthalmomyiasis, orbital and palpebral. Ophthalmomyiasis is also classified as ophthalmomyiasis externa (no eye penetration) and ophthalmomyiasis interna (penetration to inner parts of the eye occurs) (Huang *et al.*, 2020).

Calliphoridae (blow flies), Sarcophagidae (flesh flies), and *Cuterebridae* families (Diptera: Cyclorrhapha) are the causative agents of most myiasis cases (Dehghani *et al.*, 2014). According to the relationship between the host and the parasite, myiasis is divided into two types: Obligatory and facultative (Jayawardana and Yahathugoda, 2021). In obligatory myiasis, the parasite can not complete its life cycle without the host while in facultative myiasis, a host is not necessary for the parasite to complete its life cycle. Facultative myiasis (accidental) is commonly due to the oral entrance of larvae with foods (Salmanzadeh *et al.*, 2018).

Sarcophagidae (a large family with more than 2600 species) flies are ovoviviparous, depositing maggots instead of eggs on carcass, feces, and open wounds. Larval stages of *Wohlfahrtia magnifica* (Diptera: Sarcophagidae), as an obligatory myiasis agent, parasitize many warm-blooded vertebrates, which has been reported from both humans and animals such as dogs, camels, horses, leopards, and pigs, resulting in severe myiasis of mucosal membranes and wound lesions (Kheirabadi *et al.*, 2014). The myiasis caused by *W. magnifica* has been reported from different parts of Europe, Asia, and Africa (Giangaspero *et al.*, 2011). *Wohlfahrtia* sp. has been reported from various parts of Iran and different cases of human and animal myiasis due to this species have been reported previously (Akbarzadeh *et al.*, 2018; Alizadeh *et al.*, 2014; Dehghani *et al.*, 2014; Delir *et al.*, 1999; Kheirabadi *et al.*, 2014; Khoobdel *et al.*, 2013; Moshaverinia and Mehrjerdi, 2016; Nateghpour and Akbarzadeh, 2017). Around 61% of the animal traumatic myiasis in southern Iran were due to *W. magnifica* maggots and this species has been confirmed as the main traumatic myiasis agent of animals in Iran (Rafinejad *et al.*, 2014). However, no reports of orbital myiasis are reported in dogs in this country. In the present report, a case of bilateral ocular myiasis due to *W. magnifica* in an ownerless dog is described (Karaj, Iran), and a successful treatment method has been introduced.

Materials and Methods

Case presentation

In August 2021, an adult female Pitbull, weighing 15kg, was attended to a veterinary clinic, Karaj, Iran. The dog was referred to the veterinarian with severe anorexia and ocular myiasis. The lesions were seen in both eyes,

mainly in the ocular bulbs, lower eyelids, and orbital cavities (Figure 1). A close-up examination revealed the infestation with maggots, unpleasant smell, edema, erythema, ulcers, and necrotic tissues. The orbital walls of both eyes were almost not visible because of massive larvae over the eyes. The body temperature was 40.5°C representing fever.



Figure 1: Clinical manifestation of orbital myiasis in an anesthetized Pitbull dog before treatment (A and B).

After removing all larvae under complete anesthesia with Ketamine/ Xylazine, the wounds were washed with Povidone-iodine diluted in normal saline. Then, Permethrin 5% (Gilaranco Pharm Co., Iran) was applied to the wounds in order to reduce the number of live larvae and re-infestation. Oxytetracycline spray 2.5% (25mg/mL: 3.84% w/w) was applied directly to the wounded tissues and continued until the end of the second week (once daily). Systemic Antibiotic therapy with Penicillin/ Streptomycin 1+1 (Nasrpharma CO., Iran) was also used to reduce the chance of toxemia, for 5 days (10,000 units/kilogram, once daily). To reduce the inflammation, Dexamethasone was prescribed once a day for three days (0.1 mg/kg, IM). Because the dog had permanently lost its sight, food and water were given to the animal manually for three weeks.

Ethical approval

As the study was conducted with a clinical case, ethical committee approval was not required. The treatment complies with the guidelines laid down by the ethical rights of animals following the country's law and regulations.

Case follow-up

After a week of treatment, there was a progressive improvement in the animal's condition. Ophthalmic examination of both eyes revealed that the animal was not blind. As a result, permanent loss of sight which was observed at the beginning of animal presentation might be due to edema, pain, and larvae aggregating around both eyes. Examination on day 20 showed that the animal regained its vision completely. examination on day 20

showed that the animal regained its vision completely (Figure 2).

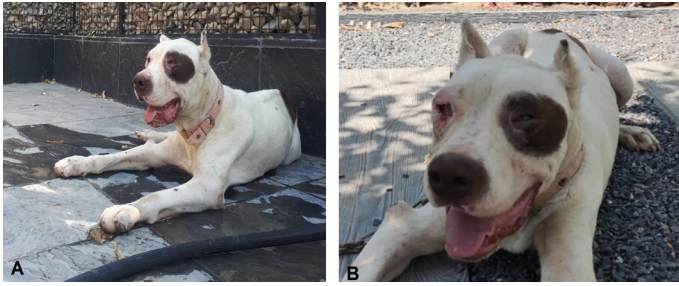


Figure 2: Improvements in animal's condition after three weeks of treatment.

Identification of larvae

larvae were removed from the wound using forceps and then kept in ethanol 70%. The collected larvae (20) were identified using relevant keys (Povolny, 1997; Szpila *et al.*, 2014; Niederegger *et al.*, 2019) using stereomicroscope (Olympus CX41, Tokyo, Japan). Larvae were cylindrical, grey in color, 14 to 18 mm in length, and 3 to 5 mm in width. They also had scattered stout spines on the body surface. The larvae of *W. magnifica* were characterized by hidden spiracles in the cavity of the posterior part. The posterior spiracle had a fairly chitinized button area with wide openings on the peritreme as well as three inner slits (Figure 3).



Figure 3: Identification of larvae by morphological keys. (A) larvae, (B) spiracles in the cavity of posterior part, (C) scattered stout spines on the body surface.

Results and Discussion

Tissue damage and economic losses (skin damages, reduction in growth rate, milk and meat production, and death due to toxemia) are subsequent outcomes of myiasis (Scholl *et al.*, 2019). The cases of myiasis have not been recorded regularly in the health system of Iran (Alizadeh *et al.*, 2014), but it seems to be prevalent in various parts especially in the areas of condensed cattle keeping industries as an occupational disease (Akbarzadeh *et al.*,

2012).

As far as we know, no report is present in Iran in terms of ocular myiasis in animals. However, many cases of myiasis in different organs due to *W. magnifica* have been reported: infestation of dogs with *W. magnifica* in different organs including mouth, paw, skin and, prepuce (Moshaverinia and Mehrjerdi, 2016), Infestation with *W. magnifica* in referred dogs with a prevalence of 1.95% in Razavi Khorasan province (Minabaji *et al.*, 2020), genital myiasis caused by *W. magnifica* in camel herds in the southwest of Iran (Kheirabadi *et al.*, 2014), and gingival myiasis in a goat caused by *W. magnifica* from Mashhad, northeast of Iran (Moshaverinia and Naiminik, 2013). In a study conducted among slaughtered animals in Kashan, 4 species of dipterous larvae including *W. magnifica* were detected on sheep (Dehghani *et al.*, 2012). Some human cases of *W. magnifica* myiasis have also been reported in Iran. Larvae were isolated from male and female external urogenital system and dental cavity (Athari and Fallah, 1993; Delir *et al.*, 1999; Salimi *et al.*, 2010).

Myiasis has been reported in human and many animal species; however orbital myiasis, especially in bilateral form, is not a common case. Miranda *et al.* reported a case of ocular myiasis of a naturally infested female Beagle dog caused by *Cochliomyia hominivorax*. The animal was prescribed orally with Nitenpyram alongside analgesic and anti-inflammatory medication. The treatment was reported to be effective (Miranda *et al.*, 2020). A case of ocular myiasis with *Lucilia sericata* in a sheep from Saudi Arabia has also been reported in 2008 (Ahdally, 2008).

Previous studies revealed Permethrin is effective on the prevention of wound re-infestation by larvae during the treatment process. Permethrin produces protection against new larval infestation for almost 2 weeks with only limited dermal absorption (<1%) (Al-Eissa *et al.*, 2008; Mohit and Shojaei, 2011). Safe permethrin products are now available for food animals in sprays and pour-on formulation to treat ectoparasites. Low dermal absorption and as a result low toxicity, long duration of action, and different available formulations made Permethrin a good drug of choice. Oxytetracycline is a protein synthesis inhibitor with broad-spectrum activity. Topical application of Oxytetracycline results in significant control of primary and secondary skin infections (Olszewska, 2006). Our treatment included Permethrin, local, as well as systemic antibiotic therapy with Oxytetracycline and Penicillin/Streptomycin, respectively and systemic anti-inflammatory dexamethasone. Case follow up to day 20 showed a significant improvement in the animal's condition including weight gain, eye vision, normal temperature, and reduced edema. Previous studies also suggested Ivermectin as a potent drug in animal myiasis cases (Al-Eissa *et al.*, 2008; Sharun and Manjusha, 2019). It has been reported

that Ivermectin is a reliable drug in larval eradication from open wounds with a long duration of action (22 days) (Al-Eissa *et al.*, 2008).

Conclusions and Recommendations

The present report revealed that mechanical removing of the larvae, subsequent washing of the wound with diluted Povidone-iodine, disinfection of the surrounding with permethrin and antibiotic therapy alongside anti-inflammatory drugs are effective in the treatment of orbital myiasis in dogs. As a result, both medical and veterinary clinicians should be aware of the possibility of the presence of myiasis in different parts of patients' bodies as well as proper treatment.

Conflict of interest

The authors have declared no conflict of interest.

References

- Ahdally, M.H., 2008. Ocular and cutaneous myiasis in sheep in Saudi Arabia. *Mansoura Vet. Med. J.*, **1**: 39–47. <https://doi.org/10.21608/mvmj.2008.125389>
- Akbarzadeh, K., Rafinejad, J., Alipour, H. and Biglarian, A., 2012. Human myiasis in Fars province, Iran. *Southeast Asian J. Trop. Med. Publ. Hlth.*, **5**: 1205.
- Akbarzadeh, K., Saghaipour, A., Jesri, N., Karami-Jooshin, M., Arzamani, K., Hazratian, T., Shabani K.R. and Afshar, A., 2018. Spatial distribution of necrophagous flies of infraorder Muscomorpha in Iran using geographical information system. *J. Med. Entomol.*, **5**: 1071–1085. <https://doi.org/10.1093/jme/tjy098>
- Al-Eissa, G.S., Gammaz, H.A., Hassan, M.F.M., Abdel-Fattah, A.M., Al-Kholany, K.M. and Halami, M.Y., 2008. Evaluation of the therapeutic and protective effects of ivermectin and permethrin in controlling of wound myiasis infestation in sheep. *Parasitol. Res.*, **2**: 379–385. <https://doi.org/10.1007/s00436-008-0983-7>
- Alizadeh, M., Mowlavi, G., Kargar, F., Nateghpour, M., Akbarzadeh, K. and Hajenorouzal-Tehrani, M., 2014. A review of myiasis in Iran and a new nosocomial case from Tehran, Iran. *J. Arthropod. Borne. Dis.*, **2**: 124.
- Athari, A. and Fallah, T., 1993. Dental cavity myiasis due to *Wohlfahrtia magnifica*. *Med. J. Islam. Repub. Iran*, **3**: 209–210.
- Dehghani, R., Sedaghat, M.M., Esmaeli, N. and Ghasemi, A., 2012. Myiasis among slaughtered animals in Kashan, Iran: Descriptive a veterinary entomological problem in the tropics. *Iran. J. Vet. Sci. Technol.*, **1**: 19–28.
- Dehghani, R., Zarghi, I. and Sayyedi, H.R., 2014. Genital myiasis of a sheep by *Wohlfahrtia magnifica*, in Ghamsar, Kashan, Iran. *Bangladesh J. Med. Sci.*, **3**: 332–335. <https://doi.org/10.3329/bjms.v13i3.15451>
- Delir, S., Handjani, F., Emad, M. and Ardehali, S., 1999. Vulvar myiasis due to *Wohlfahrtia magnifica*. *Clin. Exp. Dermatol.*, **24**: 279–280. <https://doi.org/10.1046/j.1365-2230.1999.00479.x>
- Gianguaspero, A., Traversa, D., Trentini, R., Scala, A. and Otranto, D., 2011. Traumatic myiasis by *Wohlfahrtia magnifica* in Italy. *Vet. Parasitol.*, **175**: 109–112. <https://doi.org/10.1016/j.vetpar.2010.09.028>
- Huang, Y.L., Liu, L., Liang, H., He, J., Chen, J., Liang, Q.W., Jiang, Z.W., He, J.F., Huang, M.L. and Du, Y., 2020. Orbital myiasis: A case report and literature review. *Medicine (Baltimore)*, **99**: e18879–e18879. <https://doi.org/10.1097/MD.00000000000018879>
- Jayawardana, P.P. and Yahathugoda, T.C., 2021. An accidental intestinal myiasis caused by *Cochliomyia macellaria*. *Case Rep. Pediatr.*, 2021. Article ID 6678411. pp. 6. <https://doi.org/10.1155/2021/6678411>
- Kheirabadi, K.P., Samani, A.D. and Vardanjani, H.R., 2014. A report on the genital myiasis by *Wohlfahrtia magnifica* in camel herds in southwest of Iran. *Vet. Res. Forum*, **5**: 329.
- Khoobdel, M., Akbarzadeh, K., Jafari, H., Mehrabi-Tavana, A., Izadi, M. and Mosavojazayeri, A., 2013. Diversity and abundance of medically-important flies in the Iranian triple islands; the Greater Tunb, Lesser Tunb and Abu-Musa. *Iran. J. Mil. Med.*, **14**: 327–336.
- Minabaji, A., Moshaverinia, A. and Khoshnegah, J., 2020. Frequency of ectoparasite infestation in dogs in Mashhad, Northeast Iran. *J. Vet. Res.*, **75**: 280–287.
- Miranda, F.R., Tavares, R.B., Oliveira, C.E.S., Lima, E.A.S., Campos, D.R., Scott, F.B. and Fernandes, I.J., 2020. Successful of nitempyram in treatment of ocular myiasis in orbital cavity in dog case report. *Braz. J. Vet. Med.*, **42**: e108920–e108920. <https://doi.org/10.29374/2527-2179.bjvm108920>
- Mohit, M.S. and Shojaei, S., 2011. Comparison of the two treatment protocols for wound myiasis in dogs referred to veterinary teaching hospital. *J. Vet. Clin. Res.*, **2**: 116–123.
- Moshaverinia, A. and Mehrjerdi, H.K., 2016. Canine myiasis and its causal agents in Northeastern Iran. *Iran. J. Parasitol.*, **11**: 91.
- Moshaverinia, A. and Naiminik, M.H., 2013. The report of gingival myiasis in a goat caused by *Wohlfahrtia magnifica* (Diptera: Sarcophagidae) from Mashhad, Iran. *Vet. Res. Biol. Prod.*, **26**: 26–30.
- Najjari, M., Shafiei, R. and Fakoorziba, M.R., 2014. Nosocomial myiasis with *Lucilia sericata* (Diptera: Calliphoridae) in an ICU patient in Mashhad,

- Northeastern of Iran. *Arch. Iran. Med.*, **17**: 523–525.
- Nateghpour, M. and Akbarzadeh, K., 2017. Necrophagous flies of synanthropic habitats in the South-East Iran. *Orient. Insects.*, **51**: 380–390. <https://doi.org/10.1080/00305316.2017.1314987>
- Niederegger, S., Akbarzadeh, K. and Szpila, K., 2019. Muscle attachment site patterns for species determination in West Palaearctic *Wohlfahrtia* (Diptera: Sarcophagidae) of medical and veterinary importance. *Sci. Rep.*, **9**: 1–11. <https://doi.org/10.1038/s41598-019-55127-5>
- Nouri, V. and Salehi, A., 2020. Myiasis in humans and animals. *Anim. Husband. Dairy Vet. Sci.*, **4**: 1–3 <https://doi.org/10.15761/AHDVS.1000178>
- Olszewska, M., 2006. Oxytetracycline mechanism of action and application in skin diseases. *Wiad. Lek. (Warsaw, Pol. 1960)*. **59**: 829–833.
- Povolny, D., 1997. The flesh-flies of central Europe (Insecta, Diptera, Sarcophagidae). *Spixiana Suppl.*, **24**: 1–260
- Rafinejad, J., Akbarzadeh, K., Rassi, Y., Nozari, J., Sedaghat, M.M., Hosseini, M., Alipour, H., Ranjbar, A. and Zeinali, D., 2014. Traumatic myiasis agents in Iran with introducing of new dominant species, *Wohlfahrtia magnifica* (Diptera: Sarcophagidae). *Asian Pac. J. Trop. Biomed.*, **4**: 451–455. <https://doi.org/10.12980/APJTB.4.2014C1029>
- Salimi, M., Goodarzi, D., Karimfar, M. and Edalat, H., 2010. Human Urogenital myiasis caused by *Lucilia sericata* (Diptera: Calliphoridae) and *Wohlfahrtia magnifica* (Diptera: Sarcophagidae) in Markazi Province of Iran. *Iran. J. Arthropod. Borne. Dis.*, **4**: 72–76.
- Salmanzadeh, S., Rahdar, M., Maraghi, S. and Maniavi, F., 2018. Nasal myiasis: A case report. *Iran. J. Publ. Hlth.*, **47**: 1419–1423.
- Scholl, P.J., Colwell, D.D. and Cepeda-Palacios, R., 2019. Myiasis (Muscoidea, Oestroidea). *Med. Vet. Entomol.*, **2019**: 383–419. <https://doi.org/10.1016/B978-0-12-814043-7.00019-4>
- Sharun, K. and Manjusha, K.M., 2019. Medical management of orbital myiasis using 0.1% w/v topical ivermectin preparation in a calf. *Int. J. Curr. Microbiol. appl. Sci.*, **8**: 1796–1800. <https://doi.org/10.20546/ijcmas.2019.805.208>
- Szpila, K., Hall, M.J.R., Wardhana, A.H. and Pape, T., 2014. Morphology of the first instar larva of obligatory traumatic myiasis agents (Diptera: Calliphoridae, Sarcophagidae). *Parasitol. Res.*, **113**: 1629–1640. <https://doi.org/10.1007/s00436-014-3808-x>